

# Spinning disk LEICA CSU W1:

Room: E1036 CBI

## Description

It's possible to use this Spinning disk to realize acquisition with 1 camera, 2 simultaneous cameras, Tirf, Frap, Flip,

- ❖ **Microscope** : Inverted Leica DMI8 for brightfield and fluorescence only (not DIC and Ph contrast)
- ❖ **AFC**: Adaptative Focus Control, to maintain focus for a long time
- ❖ **Spinning Head: CSU W1**
- ❖ **Atmosphere control** : temperature and Co2 control
- ❖ **Stage** : XY mot stage for multiposition recordings  
Z Piezo stage for fast z stack (Range 300µm)
- ❖ **Camera** : Evolve 512:  
512X512 pixels ; pixels size :16µm  
  
Orca Flash 4.0:  
2048X2018 pixels; pixel size: 6.5µm
- ❖ **Software** : Metamorph

### 1. Objectives:

Magnification	Objective Type	Aperture	Immersion	Coverglass	Working Distance
10X	HC PL APO	0,45	DRY	0,17	2,8mm
20X	HC PL APO CS2	0,75	Imm	Corr	0,66mm
40X	HC PL APO	1,3	oil	0,17	0,22mm
63X	HCX PL APO Lambda blue	1,4	oil	0,17	0,1mm
100X	HC PL APO	1,47	oil	0,17	0,1mm

### 2. Fluorescence filter

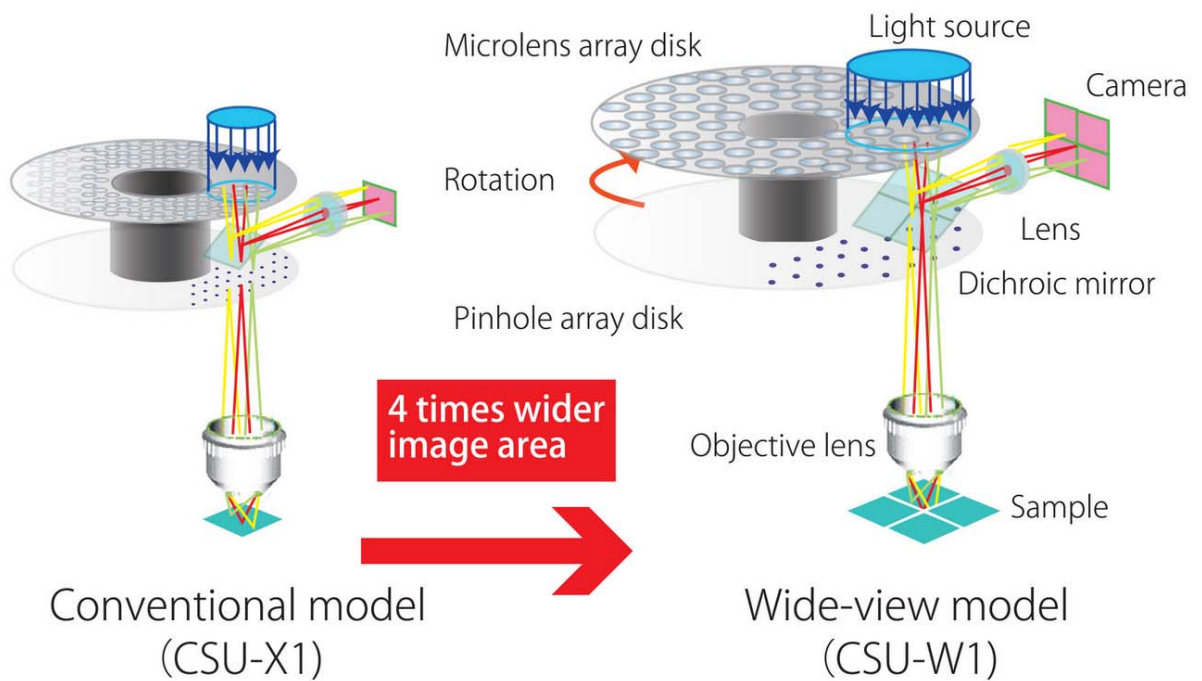
Fluorochrome	Excitation nm	Emission nm	Dichroic mirror nm
DAPI	350/50	460/50	400
CFP	436/20	480/40	455
GFP	470/40	525/50	495
TXR	560/40	630/75	585
FRAPCSU			100%
Empty	-	-	-

3. Contrast Method:

Brightfield TL	<input checked="" type="checkbox"/>	
Phase Contrasts Ph	<input type="checkbox"/>	Not possible
DIC	<input type="checkbox"/>	Not possible
Polarization	<input type="checkbox"/>	Not possible
Darkfield	<input type="checkbox"/>	Not possible
Fluorescence	<input checked="" type="checkbox"/>	DAPI, CFP, GFP TEXAS RED

4. CSU-W1:

- Principe :



- System ILAS<sup>2</sup>

The iLas MODULAR system is a unique multi-application device that offers complete control over any laser illumination. Its evolutive design allows researchers to choose and simultaneously combine :

- 360° TIRF imaging
- Single molecule imaging
- oblique illumination imaging
- FRAP
- Photo-activation
- Fast spinning disk confocal imaging
- Widefield imaging
- DUAL CAM

- Light Source

405 nm – 445 nm – 488 nm – 561 nm – 642 nm  
DAPI – CFP – GFP – Alexa 561 – Alexa 644

- Détection camera 1

1 - 452/45 → DAPI/ Hoechst...  
2 - 480/25 → CFP...  
3 - 525/50 → Alexa 488/GFP...  
4 - 609/54 → Alexa 561/ Rhodamine ...  
5 - 708/75 → Alexa 644 ...

- Detection Camera 2 ( only for dual cam)

1 - 452/45 → DAPI/ Hoechst...  
2 - 480/25 → CFP...  
3 - 525/50 → Alexa 488/GFP...

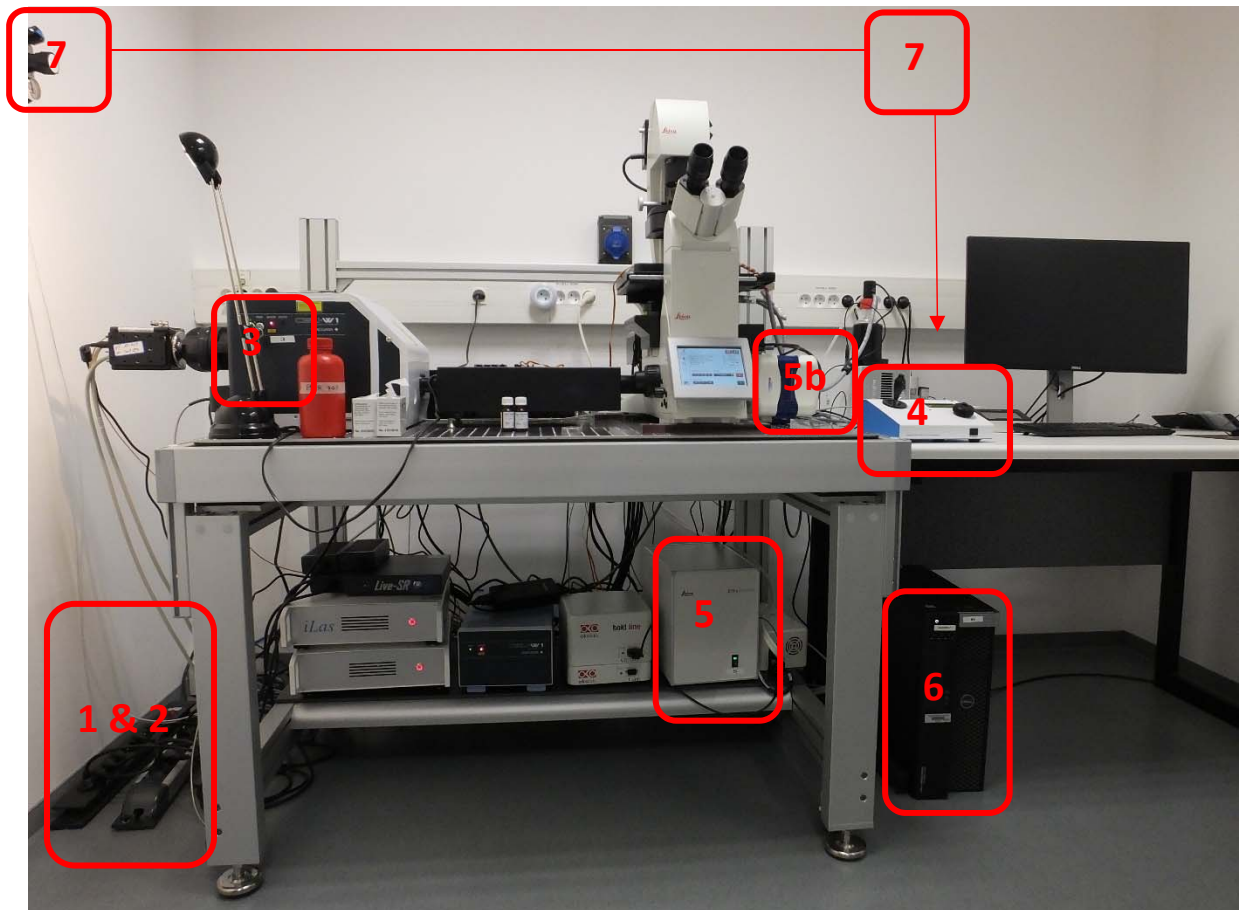
**IGBMC CBI**

**For BOOKING System :**

<http://ici-grr.u-strasbg.fr/login.php>

**For any informations :** [groupe-mic-photon@igbmc.fr](mailto:groupe-mic-photon@igbmc.fr)

## Procedures of starting spinning disk



1 & 2 – Turn on the system (multiple plug X2 )

3 – turn the key of the spinning head (CSU W1) on the position « ON »

4 – Switch on the stage controller

5 – Switch on the Microscope controller

5b – Switch on the EMCCD camera if you use the TIRF system

6 – Turn on the computer

7 – If you use the atmosphere control, turn On the Co<sup>2</sup> Valve und the multiple plug

# The Microscope: Leica DMI 8

## Control panel field on the left microscope side

- 1 Motorized aperture diaphragm adjustment
- 2 Motorized illuminated field diaphragm adjustment
- 3 Motorized toggling between the TL/Fluor illumination axes
- 4 Adjusting the Transmitted Light brightness of the intensity levels for the automated FIM (fluorescence intensity manager)

An LED indicates the illumination method which is currently affected by the brightness adjustment.

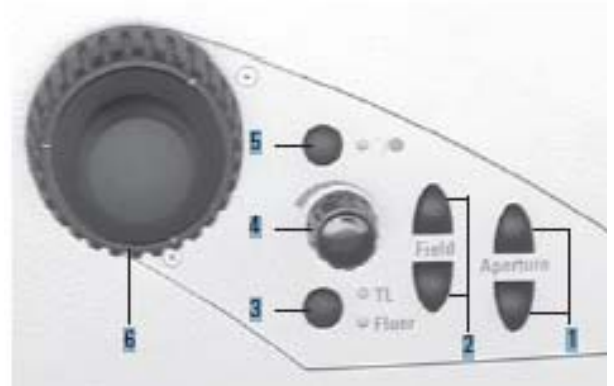
- 5 Opening/closing the motorized shutter or during manual shutter light control

An LED displays the state of the shutter:

LED on = shutter opened

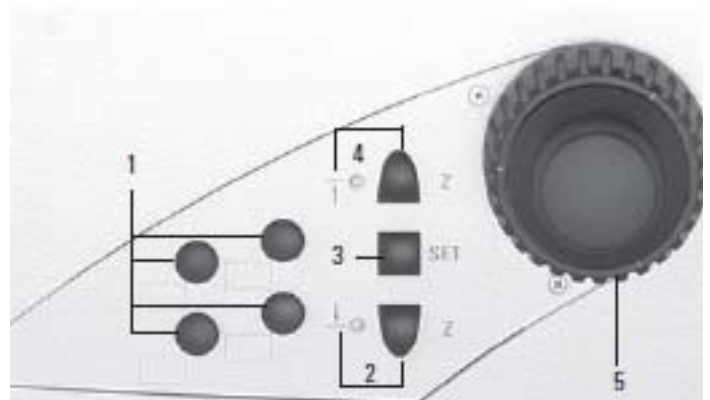
LED off = shutter closed.

- 6 Focusing



## Control panel field on the right microscope side

- 1 Function Keys to move the objective turret and filter Cube turret
- 2 Focus threshold (the LED illuminates when the focus threshold is set)
- 3 Setting the focus position and focus threshold
- 4 Focus position (the LED illuminates when the focus position is set)
- 5 Focusing

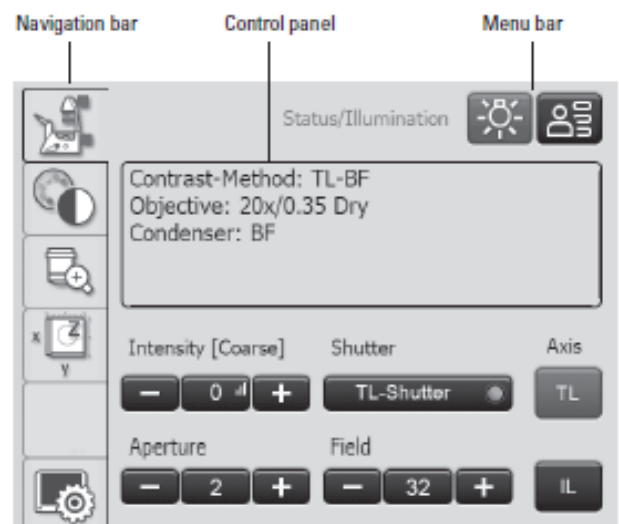


## The Touch Screen

The Touch Screen adjusts all motorized components of the Leica DMI8 automatically via buttons.

After the device is switched on, the display shows the current microscope status.

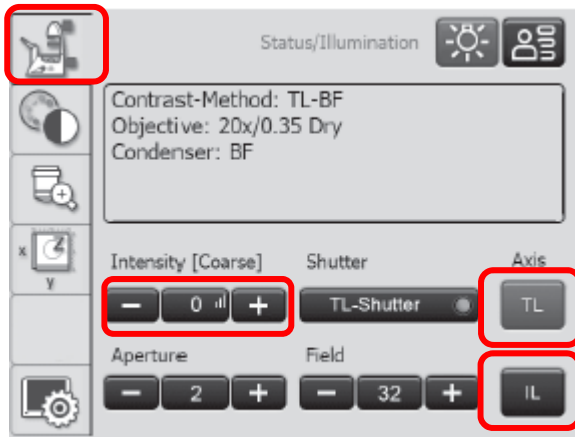
The display and functions that can be operated using the Touch Screen depend on the features of the individual microscope.



Touch Screen after initialization



### Basic microscope settings

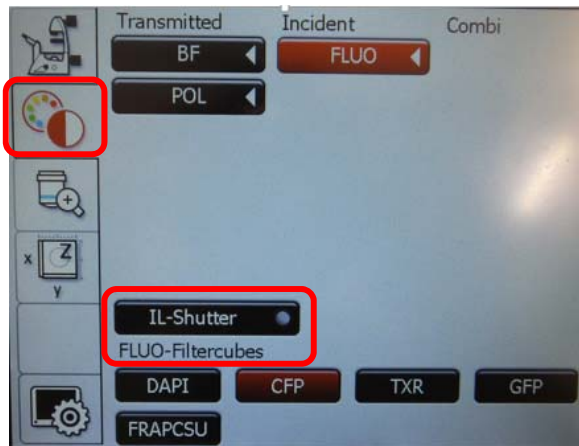


Adjusting the Transmitted Light Brightness

Adjusting the Fluorescence Light Brightness



### Contrast methods



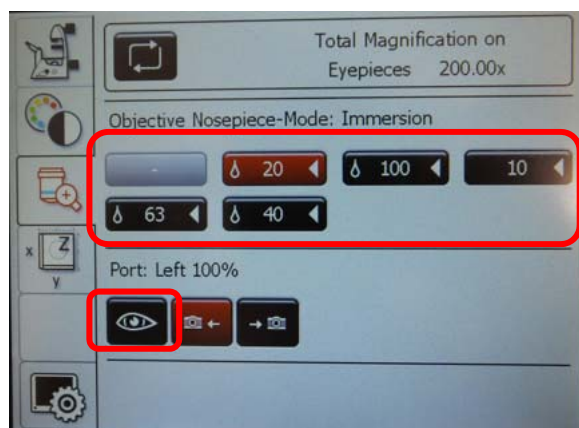
Select the contrast method

Select the filter cube if it's necessary

Switch on the lamp with the shutter button



### Magnification

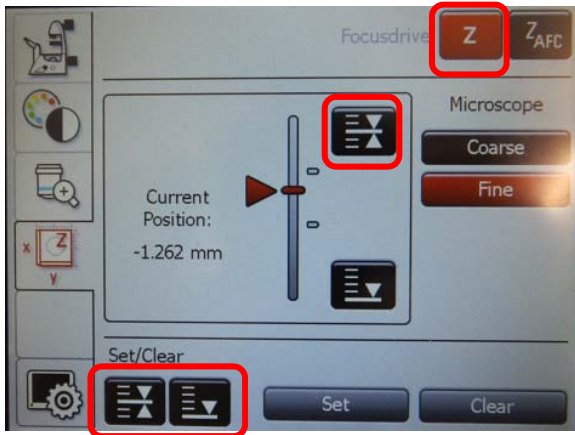


Select the magnification (all objectives are immersion oil except the **10X is dry**)

Select the right or left side port using the corresponding key or point 100% of the light at the eyepiece.



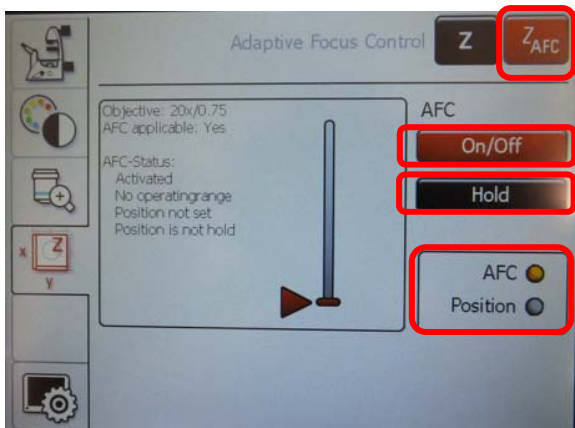
### Stage and focus controls



Select the **Focus Drive** menu using the key. The current Z position is shown in the information field.

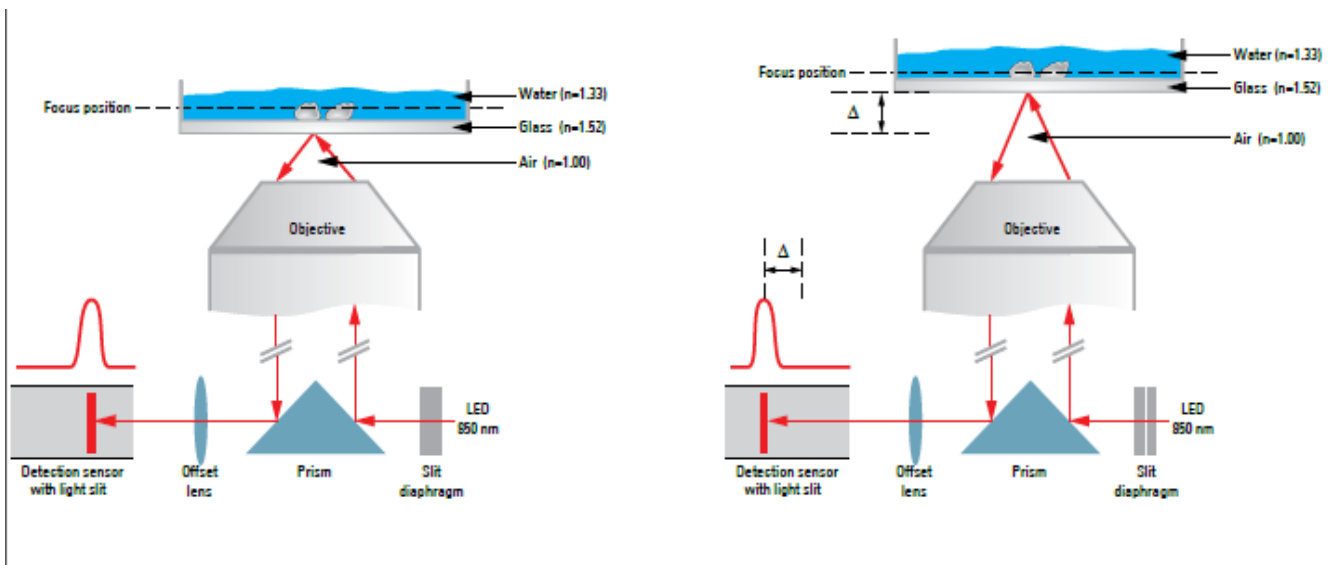
When the focus position is set, it can be approached directly using the key. To set or clear the focus position or the lower threshold, select the corresponding position under **Set/Delete**, then press the **Set** or **Delete** key.

### Leica Adaptive Focus Control



The AFC can be used to actively hold a defined focus position and automatically readjust this position regularly. This is useful if, for example, a temperature change is caused by opening the climate chamber during living cell experiments at 37°C. The AFC is activated, then the specimen on the stand is focused using the handwheel and the stopping position is saved.

Activate the AFC using the **On/Off** key. The knob behind **AFC** turns green. Focus the specimen and save the position using the **Hold** key. If the position is held, the knob behind **Position** will turn green.



# Software: METAMORPH



Double clics on your METAMORPH ICON

The screenshot shows the MetaMorph software interface. The top menu bar includes File, Edit, Regions, Stack, Acquire, Devices, Display, Process, Log, Measure, Journal, Apps, Window, and Help. The toolbar contains various icons, with the 'MDA' icon highlighted by a red box. The left sidebar contains a list of icons, with three groups highlighted by red boxes: 1) A group of five 'EVE' icons (EVE Trans, Set Shutter, EVE DAPI, EVE CFP, EVE GFP, EVE mCherry) with a callout box stating 'Short Cut to drive the contrast method of microscope'. 2) A group of five magnification icons (10x, 20X IMM, 40X OIL, 63X OIL, 100XTirf OIL) with a callout box stating 'Shortcut to drive the magnification microscope'. 3) A group of four camera icons (CSU CAM1, CSU DUAL CAM, TIRF EVOLVE, CSU FRAP) with a callout box stating 'Shortcut to choose the software configuration'. The 'MDA' icon in the toolbar is also highlighted with a red box and a callout box stating 'Shortcut to open the Multi-Dimensional Acquisition'.

Short Cut to drive the contrast method of microscope

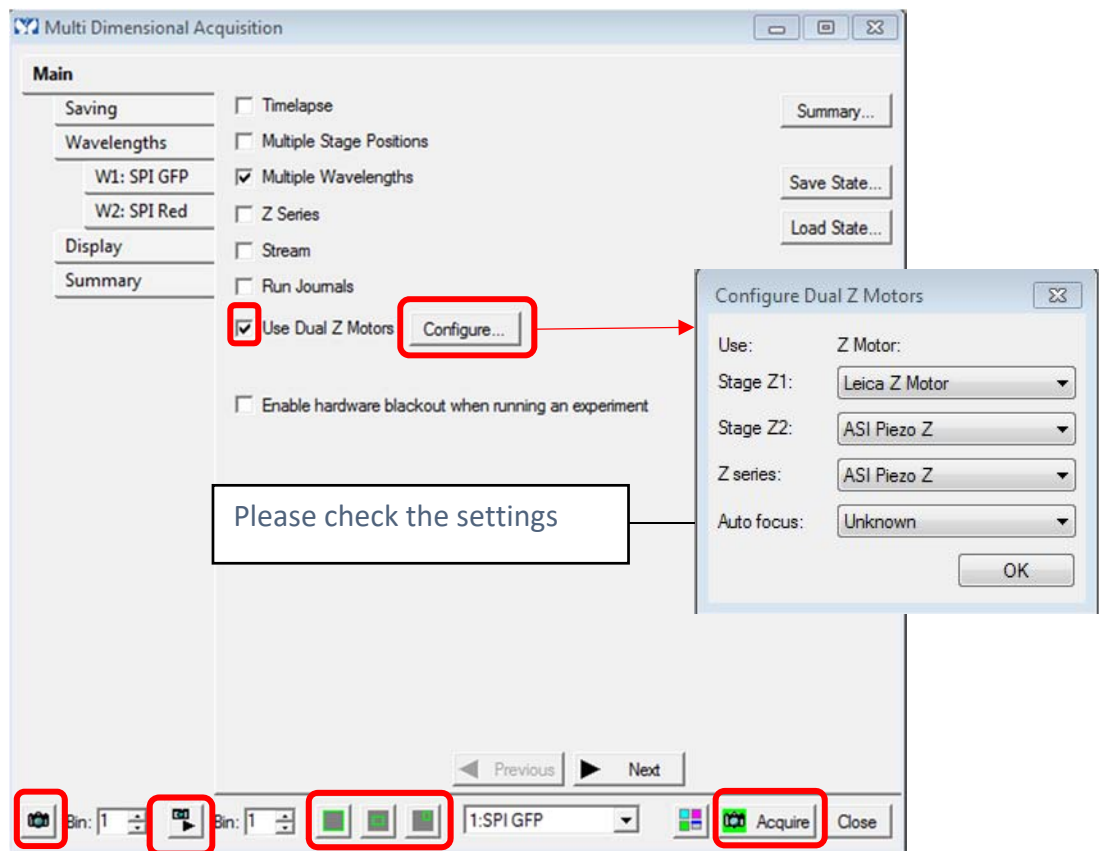
Shortcut to drive the magnification microscope






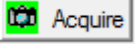
Shortcut to choose the software configuration

Shortcut to open the Multi-Dimensional Acquisition

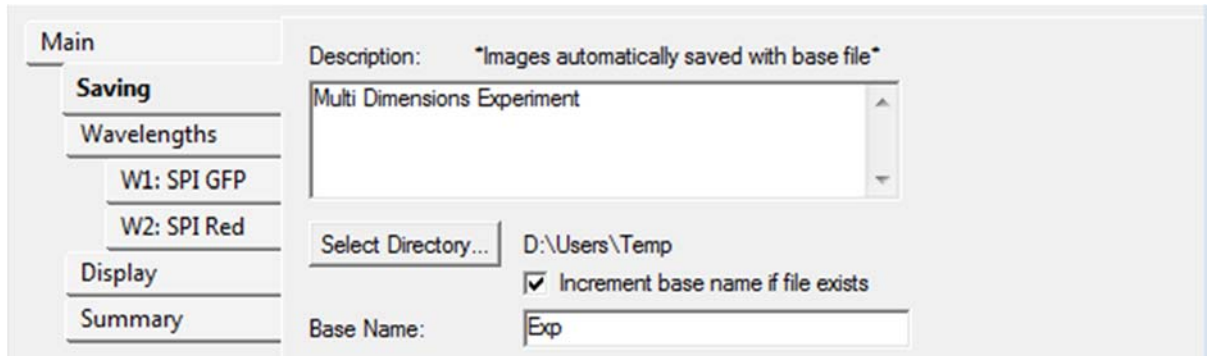


1 – Clic on MDA Icon, a new windows opens. In this interface, you can choose what you want to do.



	SNAP	Acquire an image using the current settings specified in the Acquire main dialog box
	LIVE	Showing a live image
	FULL CHIP	Initialiez the full chip of your camera 512x512
	Center CHIP	To use center chip of your camera 256x256
	ROI CHIP	To use only one ROI chip of your camera
	ACQUIRE	Start MDA experiment

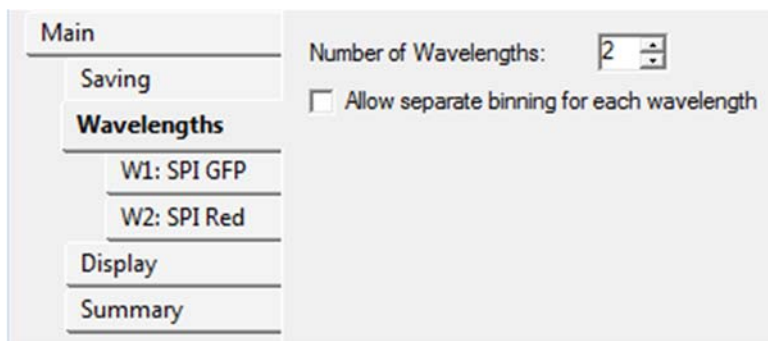
### 1.1 Saving :



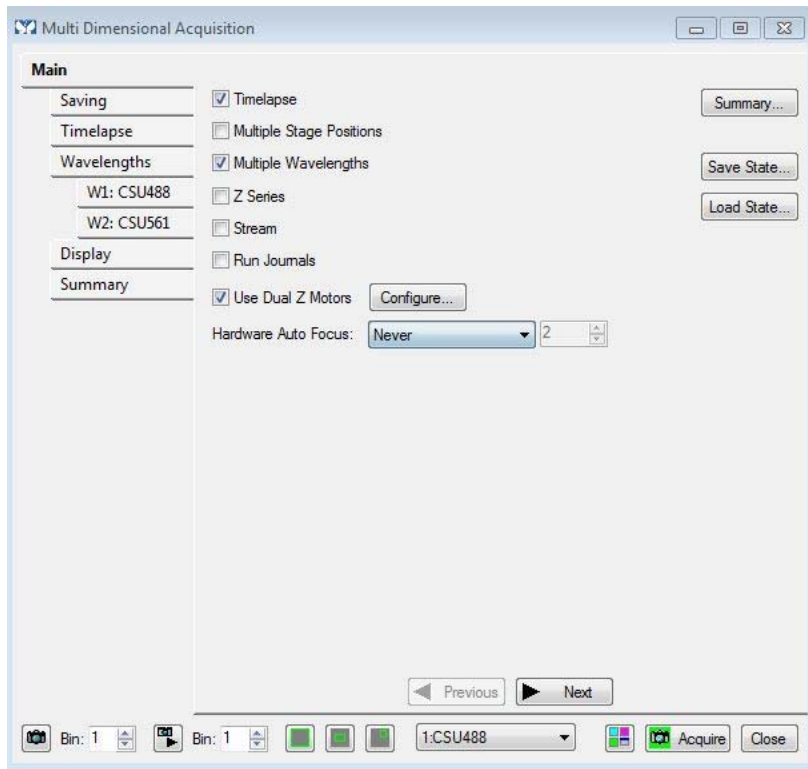
In this window, select directly the Save folder by clicking on the Select directory icon, it's possible to use the base name and the software will automatically increment the number of the image.

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### 1.2 Multiple Wavelengths :

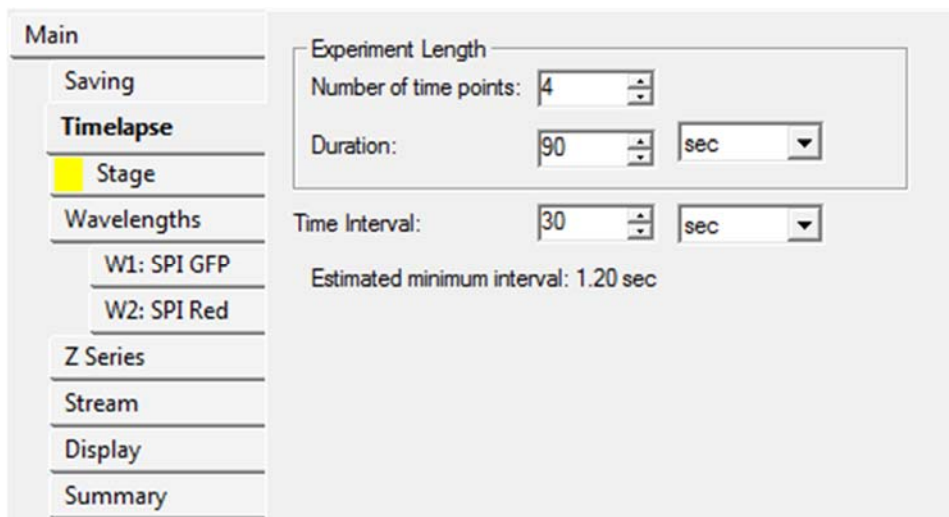


In this window you select the number of channel (wavelengths).



- After select the illumination you want to use
- You adjust the exposure time (you open the live image for parameter this value)

### 1.3 Timelapse

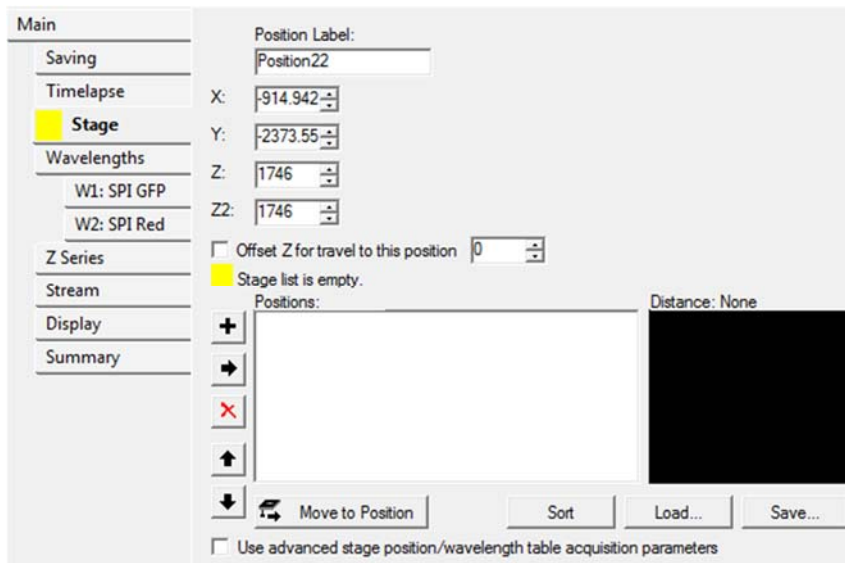


Enter your acquisition parameters.

Be careful, the « Estimated minimum interval » is wrong value. The system doesn't consider mechanical movements.

Please test your timelapse with 2 timepoints for estimate the good minimum interval.

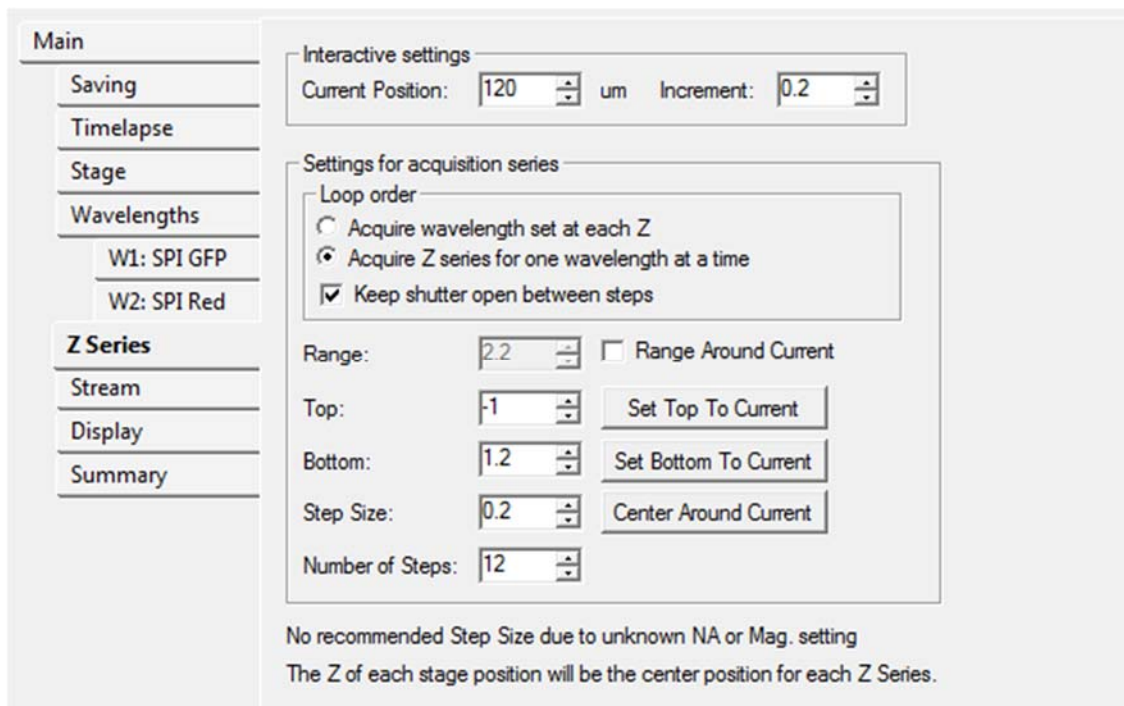
## 1.4 Stage



Select your sample area and clic on **+** bouton, the software consider the x,y and z position.

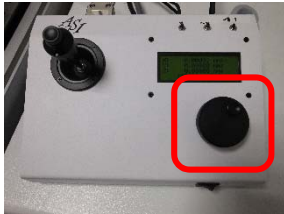
You can name diferents positions on Position Label, the software increase automaticly.

## 1.5 Z Series



There are two modes for the z stack acquisition

- The **Top and Bottom** mode : for this mode you open the live image and you change the z position with the knob of the stage controller or the cursor “Current Position”   on the Interactive settings windows.



- The second mode is if you know the thickness of your sample (or it's possible to estimate by the « top-bottom » mode) you check the « Range Around Current » box and you indicate the range value. This mode is recommended if you want to combine multipositions and z series.

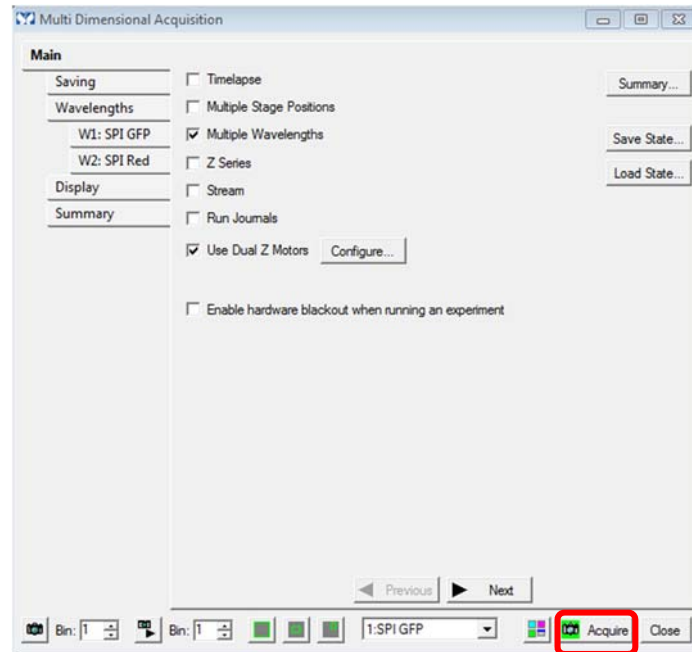
### 1.6 stream : Rapidly acquires

The screenshot shows the 'Stream' configuration window. On the left is a sidebar with tabs: Main, Saving, Timelapse, Stage, Wavelengths (with sub-tabs W1: SPI GFP and W2: SPI Red), Z Series, **Stream**, Display, and Summary. The main panel has the following settings:

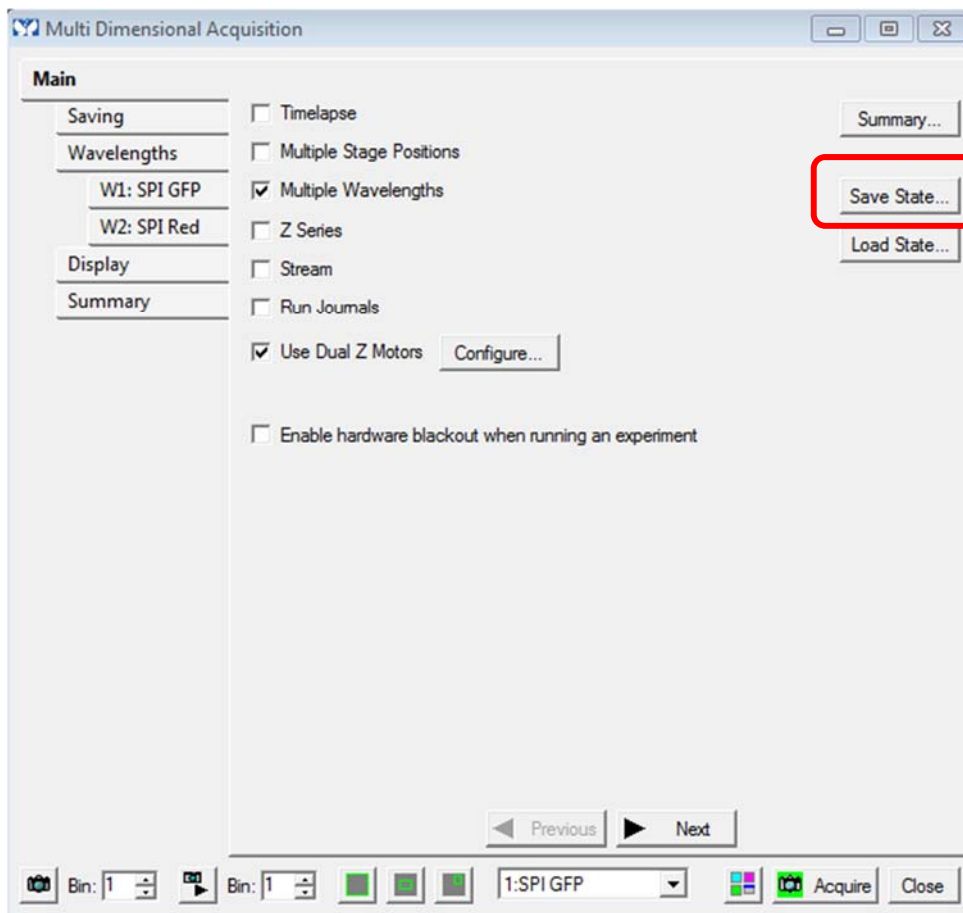
- Stream Time
- Stream Z
- Stream Multiple Wavelengths
- User Program Name: [None]
- Frames to skip when resetting Z: 0
- Stream Intensifier Gain: 0
- Stream Exposure Time (ms): 50
- Display preview during acquisition
- Update preview every 1 frames
- Enable Z Remove
- Status: ■ Stream configuration OK
- Stream To: Hard Disk
- Start Frame: 0
- Memory Required 12.00 MB
- Memory Available 0.00 KB

Select what you want to do and set in the different tabs (Timeapse, Zseries, Wavelengths).

## 2- Acquire:

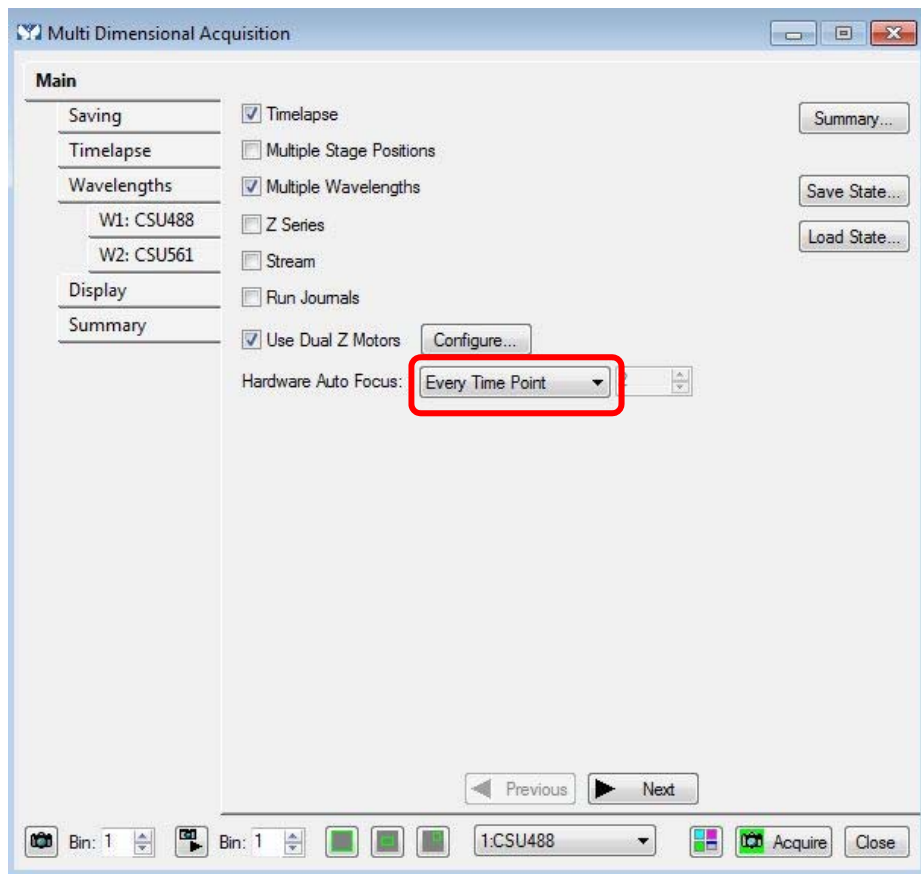


When the settings are completed, clic on the acquire Icon



It's possible to save the complet experiment for the next session by the "Save state" Icon.

## 1.7 AFC to maintain the Focus during a time-lapse acquisition:



When you set your experiment, if you select « Timelapse », there is a new part on your « Main » menu : « Hardware Auto Focus ». there are different options but in the majority of cases, « Every Time Point » is a good compromise.

If « Never » is selected, the AFC is disabled.